

# Resistance against strongylid nematodes in two high prevalence Equine Recurrent Airway Obstruction families has a genetic basis

Päivi Nussbaumer Schleuniger<sup>1</sup>, Caroline F. Frey<sup>2</sup>, Bruno Gottstein<sup>2</sup>, June E. Swinburne<sup>4</sup>, Gaudenz Dolf<sup>3</sup> and Vinzenz Gerber<sup>1</sup>

Equine Clinic, Department of Veterinary Clinical Studies<sup>1</sup>, Institute of Parasitology, Department of Paraclinics<sup>2</sup> and Institute of Genetics, Department of Clinical Research & VPH<sup>3</sup>, Vetsuisse-Faculty, University of Berne, Switzerland and Animal Health Trust, Newmarket, Suffolk U.K.<sup>4</sup>

## Summary

A recent study showed increased resistance against strongylid nematodes in offspring of a stallion affected by recurrent airway obstruction (RAO) compared with unrelated pasture mates. Resistance against strongylid nematodes was associated with RAO affection. Hypothesis: Resistance against strongylid nematodes has a genetic basis. The genetic variants influencing strongylid resistance also influence RAO susceptibility. Faecal samples from the half-sibling offspring of two RAO-affected Warmblood stallions – 98 offspring from the first family (family 1) and 79 from the second family (family 2) – were analysed using a combined sedimentation-flotation method. The phenotype was defined as a binary trait - either positive or negative for egg shedding. The influence of non-genetic factors on egg shedding was analysed using SAS, the mode of inheritance was investigated using PAP and iBay, and the association between shedding of strongyle eggs and RAO was estimated by odds ratios. Previously established genotypes for 315 microsatellite markers were used for QTL analyses using GRID QTL. The inheritance of “strongylid egg shedding” is influenced by major genes on ECA15 and ECA20. Shedding of strongylid eggs is associated with RAO in family 1 but not in family 2. Conclusions: The status of “shedding of strongyle eggs” has a genetic background. The results were inconclusive as to whether “egg shedding” and RAO share common genetic components. Our results suggest that it may be possible to select for resistance against strongylid nematodes.

**Keywords:** horse / strongylid nematodes / recurrent airway obstruction / resistance / major gene

---

## Resistenz gegen kleine Strongyliden in zwei Warmblut-Familien mit erhöhter Prävalenz rezidivierender Atemwegsobstruktion hat eine genetische Grundlage

Eine kürzlich erschienene Arbeit zeigte eine erhöhte Resistenz gegen kleine Strongyliden bei Nachkommen eines Hengstes mit rezidivierender Atemwegsobstruktion (Recurrent Airway Obstruction (RAO)) im Vergleich mit ihren nichtverwandten Weidekameraden. Die Resistenz gegen kleine Strongyliden war mit RAO assoziiert. Hypothese: Resistenz gegen kleine Strongyliden hat eine genetische Basis. Die genetischen Varianten welche die Resistenz gegen kleine Strongyliden beeinflussen, beeinflussen auch die RAO-Anfälligkeit. Material und Methoden: Kotproben von Nachkommen zweier RAO-betroffenen Warmbluthengste – 98 Nachkommen der ersten Familie (Familie 1) und 79 Nachkommen der zweiten Familie (Familie 2) – wurden mittels kombinierter Sedimentations-Flotations-Verfahren untersucht. Der Phänotyp wurde als binäres Merkmal definiert, entweder positiv oder negativ für die Eiausscheidung. Der Einfluss von nicht-genetischen Faktoren wurde mittels SAS analysiert, der Erbgang wurde mittels PAP und iBay untersucht und die Assoziation zwischen Ausscheidung von Strongylideneier und RAO wurde mittels Odds Ratio geschätzt. Die früher determinierten Genotypen für 315 Mikrosatelliten wurden für die QTL Analyse (GRID QTL) gebraucht. Die Vererbung der Ausscheidung von Eiern kleiner Strongyliden wird durch Gene auf ECA15 und ECA20 beeinflusst. Die Ausscheidung von Eiern kleiner Strongyliden ist mit RAO in Familie 1 assoziiert aber nicht in Familie 2. Der Status “Ausscheidung von Strongylideneier” hat einen genetischen Hintergrund. Die Resultate sind als ersten Hinweis auf einen gemeinsamen genetischen Hintergrund für Eiausscheidung und RAO zu werten. Unsere Resultate deuten darauf hin, dass es möglich sein könnte für eine Resistenz gegen kleine Strongyliden zu selektionieren.

**Schlüsselwörter:** Pferd / kleine Strongyliden / rezidivierende Atemwegsobstruktion / Resistenz / Gen

## Introduction

Since the hygiene hypothesis was postulated by *Strachan* (1989), the investigative interest in associations between parasite, especially helminth infection and development of hypersensitivities has greatly increased. The most common hypersensitivity-associated disease in horses is recurrent airway obstruction (RAO). RAO shares many characteristic features with human asthma (*Robinson* 2001): e.g. airway inflammation, mucus accumulation, reversible airway obstruction and bronchial hyperresponsiveness (*Martin* 2001, *Gerber* et al. 2004).

RAO is strongly influenced by environmental factors, mainly allergens in hay dust, but a genetic predisposition for this

disease has also been demonstrated (*Marti* et al. 1991, *Ramseyer* et al. 2007, *Gerber* et al. 2009). The prevalence and clinical manifestation of RAO was found to be increased in two large half-sib sire families (referred to here as family 1 and family 2) in comparison with a control group (*Ramseyer* et al. 2007). In family 1, *Jost* and co-workers (2007) demonstrated association and linkage of RAO with microsatellite markers on ECA13 near the interleukin 4 receptor chain (IL4R) gene. Polymorphism in human IL4R are associated with phenotypes related to asthma and atopy (*Hershey* et al. 1997, *Ober* et al. 2000). Moreover, IL4R is associated with parasitic defence in humans and animals (*Scales* et al. 2007). *Neuhaus* and co-workers (2008) observed that offspring of the RAO-affected sire with

the IL4R association (family 1) were more resistant against strongylid nematodes compared to a matched control group. Helminthiasis seems to play a beneficial, even protective role in the context of autoimmune diseases and allergies (da Rocha Fa 2006, McKay 2006). The classical hygiene hypothesis postulates that infections and unhygienic contact may confer protection against the development of allergic diseases. Another possible explanation for the inverse relationship between resistance to certain parasitic diseases and asthma is that genetic determinants which confer protection against detrimental worm burdens are the same determinants involved in atopic asthma (Barnes et al. 2005). This alternative hypothesis could also explain the observation that individuals with a history of atopy appear to be more resistant to helminthic parasites (Lynch et al. 1998).

The family of strongylids contains both the large strongyles (Strongylinae) which are nowadays rarely found in horses in Switzerland (Meier and Hertzberg 2005) and the small strongyles (Cyathostominae) which are the most important parasitic pathogens of grazing horses (Love and Duncan 1992). Although many horses harbour large burdens of cyathostomes they rarely develop clinical signs (Love et al. 1999).

Neuhaus and co-workers (2008) observed that offspring from family 1 had a decreased risk of severe infestation with strongylids as compared to unrelated, RAO-unaffected pasture mates, matched for age, breed, stable, pasture and deworming management. Furthermore, within the offspring from family 1, the RAO-affected individuals had a >5-fold reduction in shedding of parasite eggs compared to unaffected offspring. This raised the question whether there is a relationship between resistance to helminth infections and susceptibility to RAO in horses in general. An investigation in family 2 found no significant difference in parasite infestation level between the offspring and the matched control group (Bründler et al. 2011). However, Bründler et al. (2011) did observe that unrelated RAO-affected Warmblood horses shed fewer strongylid eggs than their matched controls.

To our knowledge, a possible genetic basis for the status of patent nematode infection and respective "egg shedding" in horses has not yet been investigated. Therefore, the main goal of this study was to elucidate the role of genetics in the status of "egg shedding" in family 1 and 2 by investigating the mode of inheritance and performing genetic association and linkage analysis with microsatellite markers. In addition, we investigated whether "parasite egg shedding" and RAO share a common genetic basis in family 1 and 2.

## Materials and Methods

### Animals

The study was approved by the Animal Experimentation Committee of the Canton of Berne, Switzerland (authorisation number BE 33/07). Two half-sibling Warmblood families have been investigated with respect to RAO (Ramseyer et al. 2007). On the basis of a standardised questionnaire, the RAO status of all horses was classified using the Horse Owner Assessed Respiratory Signs Index (HOARSI), as described in detail by Ramseyer and co-workers (2007). Briefly, owner

supplied information on coughing, nasal discharge, performance and breathing pattern was used to assign the HOARSI classification which has four grades: unaffected (1), mild (2), moderate (3) and severe (4) clinical signs. The classification refers to the period when the horses showed the most severe clinical signs.

In the present study, 98 offspring from family 1 and 79 offspring from family 2 were included. Family 1 comprised 43 females and 55 males ranging in age from 9 to 17 years with an average of 13.5 years, while family 2 comprised 46 females and 33 males ranging in age from 7 to 15 years with an average of 12.5 years. Thirty-four (35%) descendants of family 1 were considered unaffected (HOARSI 1), 21 (21%) were classified as HOARSI 2, 29 (30%) as HOARSI 3 and 14 (14%) as HOARSI 4. Nineteen (24%) descendants of family 2 were considered unaffected (HOARSI 1), 25 (32%) were classified as HOARSI 2, 25 (32%) as HOARSI 3 and 10 (12%) as HOARSI 4. All 177 individuals had been previously genotyped with 315 microsatellite markers (Swinburne et al. 2009).

### Faecal sample collection

A sample was collected from all 177 horses and information was gathered through a questionnaire. The horses were located in 160 stables. All owners were asked what anthelmintic treatment specifically was used as the last deworming treatment. Further, information was collected on pasture management, such as removal of manure from pasture, other species on the pasture, or mowing of the pasture, geographic region and altitude. The faecal samples were only collected after the egg reappearance period (ERP), defined as the period when horses are expected to reach a maximum positive egg count after treatment (von Samson-Himmelstjerna 2006). Since different anthelmintics were used by different owners, a maximum egg reappearance period of 90 days was chosen to include long-active agents such as Moxidectin (von Samson-Himmelstjerna 2006). Horses were excluded when this protocol was not adhered to. Samples collected from the stable floor had to be fresh and uncontaminated by other faeces or mud. If no faeces were readily available, the faecal samples were recovered rectally.

### Coprology

Each sample was examined by only one person without knowledge of the health status of the horses and the family they belonged to. Samples were analysed separately by a combined sedimentation-flotation method (Bauer 2006). The results were expressed as a binary trait: "egg shedding", positive or negative.

### Statistics

The four HOARSI categories were combined into two classes, where HOARSI 1 was considered unaffected and HOARSI 2, 3 and 4 affected. A possible association between HOARSI and "egg shedding" was investigated by calculating odds ratios (<http://statpages.org/ctab2x2.html>)<sup>1</sup>. For the joint ana-

lysis of both families PROC GLIMMIX of the SAS package<sup>2</sup> was used to estimate non-genetic effects on the probability of egg shedding in a linear model. The effects entering the model were sire, gender, HOARSI, age, removal of manure from pasture, geographic region, altitude ( $\leq 500\text{m}$  and  $>500\text{m}$  above sea level), sharing pasture with other species (livestock) and mowing of the pasture. Segregation analyses were performed using PAP<sup>3</sup> together with NPSOL<sup>4</sup> and iBay1.44<sup>5</sup>. Linkage between egg shedding and the microsatellite markers were calculated using GRID QTL (<http://www.gridqtl.org.uk/> Seaton et al. 2006). A single QTL model with mowing as a fixed effect was fitted at 1 Mb steps along the chromosomes. A QTL was considered present if the F statistic exceeded a chromosome-wide significance level of  $P=0.05$ . All analyses were performed with each family separately and also with both families combined. F statistics were also compared with a genome-wide significance level at  $P=0.05$ .

## Results

### *Coprology; association between "egg shedding" and HOARSI*

In the majority of the faecal samples, no "egg shedding" was detected. The percentage of offspring "shedding eggs" was significantly lower ( $P<0.01$ ) in family 1 (12%) than in family 2 (34%). Shedding of strongylid eggs and HOARSI were associated in family 1 but not in family 2 (Table 1).

### *Non-genetic effects on "egg shedding"*

The effects on "egg shedding" of gender, HOARSI, age, removal of manure from pasture, geographic region, altitude, sharing pasture with other species, and mowing of the pasture were not significant ( $P>0.05$ ). However, the effect of the sire on "egg shedding" was highly significant ( $P=0.01$ ). Mowing of the pasture became significant ( $P=0.02$ ) when sharing pasture with other species HOARSI, geographic region and altitude were all removed from the model. After removing all non-significant effects the final model comprised only sire ( $P<0.01$ ) and mowing of the pasture ( $P=0.02$ ).

**Table 1** Odds ratios (OR), confidence intervals (CI) and two-sided P-values for the association between presence of eggs and HOARSI in the two half-sibling sire families. The percentage of the offspring shedding eggs is given in the column ES for the two families.

	ES%	OR	95% CI	$P_{2\text{-sided}}$
Family 1	12	0.257	0.074-0.901	0.045
Family 2	34	1.235	0.413-3.665	0.785

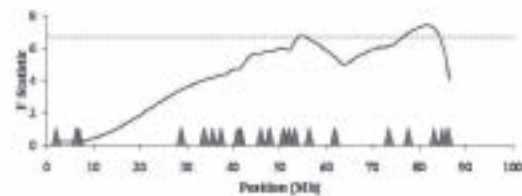
**Table 2** Parameter estimates for the separate and joint analyses of families 1 and 2

	Family 1 & Family 2	Family 1	Family 2
Frequency of allele a	0.50	0.54	0.50
Penetrance for AA	0.03	0.01	0.11
Penetrance for Aa	0.14	0.10	0.33
Penetrance for aa	0.42	0.45	0.64
Major gene <sup>1</sup>	0.48	0.50	0.33
Polygenic component <sup>2</sup>	0.39	0.42	0.60

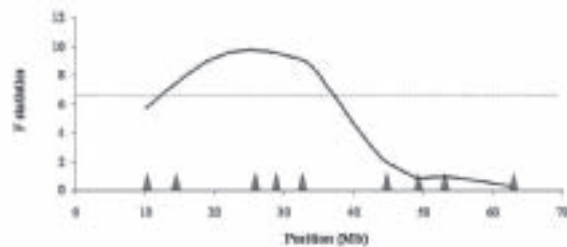
<sup>1</sup> Part of the total variance explained by the major gene, <sup>2</sup> Part of the total variance explained by the polygenic component

## *Segregation analyses*

Segregation analyses were performed for both sire families jointly and for each family separately. Using PAP with NPSOL, in all three analyses a general genetic model explained our data much better than an environmental model ( $P<0.01$ ). In all three cases the general genetic model could not be differentiated from the mixed inheritance model ( $P>0.05$ ). Also the major gene model and the polygenic model could not be differentiated from the mixed inheritance model ( $P>0.05$ ). Using iBay mixed inheritance models fitted the data in all three cases better, with higher Bayes factors (BF), than the other models. Parameter estimates were averaged over four to seven runs with 10 chains per run. The estimates of the frequency of the deleterious allele "a" and the penetrances are given in Table 2, together with the percentages of the total



**Fig 1** F statistics of the QTL analyses on ECA15 in family 1. The broken horizontal line indicates the chromosome-wide significance threshold at  $P<0.05$ . The positions of informative markers are shown with grey triangles.



**Fig 2** F statistics of the QTL analyses on ECA20 in family 2. The broken horizontal line indicates the chromosome-wide significance threshold at  $P<0.05$ . The positions of informative markers are shown with grey triangles.

variance explained by the major gene and the polygenic component.

### QTL analyses

QTL analyses using mowing as a fixed effect resulted in significant signals ( $P < 0.05$ , chromosome wide) on ECA15 (Fig 1) for family 1 and on ECA20 (Fig 2) for family 2. The joint analysis of both families gave a significant signal on ECA20 but not on ECA15 and revealed an additional signal on ECA11 ( $P < 0.05$ ). None of the signals reached genome wide significance.

### Discussion

Our results suggest the presence of more than one major gene influencing strongylid egg shedding in horses. Regions harbouring one or more major genes for egg shedding were identified on ECA15 in family 1 and on ECA20 in family 2. Egg shedding was not only influenced by two different chromosome regions in the two families, but it was significantly less frequent in family 1 than in family 2. In addition, egg shedding was significantly associated with RAO in family 1 but not in family 2. Egg shedding among the offspring in family 2 (with 34% positive faecal samples) was almost three times as frequent as in family 1 (12%). This disparity could be explained by different frequencies of the unfavourable allele of the putative major genes which are located on different chromosomes in the two families. The association of egg shedding with RAO in family 1 could also be an artefact as the two families were collected to investigate RAO and therefore are not random samples of the population. Other explanations for this association such as linkage between egg shedding and RAO or pleiotropy were evaluated but our data did not support one hypothesis or another. A larger, random sample would be needed to further investigate these hypotheses.

Our investigation was a field study in which natural infestation was examined, rather than a controlled clinical study in which each horse would be exposed to an equal infection pressure. However, post-mortem surveys of naturally occurring gastrointestinal cyathostome burdens of horses have indicated that virtually all grazing horses are infected (Ogbourne 1976, Reinemeyer et al. 1984, Krecek et al. 1989). For this reason, we assumed that the horses in our study were exposed to a similar infection pressure. Geographic region and altitude had no effect on egg shedding supporting our assumption of a similar infection pressure across Switzerland. Gender also had no effect as the sex ratio did not significantly differ in the two groups in which eggs were present or absent. As expected in a study of horses aged over 7 years, age had no effect. High faecal egg counts and clinical cyathostomiasis are more frequently seen in young horses (Love and Duncan 1992, Giles et al. 1985). HOARSI had no apparent effect on egg shedding status, which may be due to the combined analysis of families 1 and 2. The significant effect of mowing on egg shedding could be explained by the fact that mowing reduces the number of infectious strongylid larvae on the grass (Ardeleanu et al. 2008). The sire had by far the largest effect on the probability of egg shedding, clearly indicating a genetic component. The large effect of the sire could possibly obscure the effects of other factors.

In both families, the segregation analyses showed the presence of a major gene. However, the results also suggested that the gene influencing egg shedding status is not the same in the two families as the penetrance and the percentage of the variance explained by the major gene differed. Furthermore there must be other genes associated with egg shedding besides these major genes as the polygenic component explained a considerable part of the variance in both families (Table 2).

The separate QTL analysis of both families gave a significant chromosome wide signal on ECA 15 for family 1 and on ECA20 for family 2. This result suggests that for egg shedding, as for RAO (Gerber et al. 2009), different major genes play a role in the two families. However, the signal on ECA11 suggests an overlap of the genetic background in the two families.

There are several candidate genes on ECA15, such as the interleukin 1 receptor genes or the interleukin 18 receptor accessory protein gene, all involved in the development of asthma (Nakae et al. 2003, Ishikawa et al. 2006) and in the defence against parasites (Titus et al. 1991, Sasaki et al. 2005).

The genes of the major histocompatibility complex (MHC) class I, II and III in horses are located on ECA20. The MHC plays an important role in the pathogenesis of allergic diseases (Blaser 2008) and in the defence against helminths (Paterson et al. 1998). The MHC also has been implicated with nematode resistance in sheep (Davies et al. 2006).

In conclusion, our results point at the presence of major genes on ECA15 and ECA20 affecting strongylid egg shedding and possibly RAO as well. This needs to be confirmed in the sample of the general equine population. It should be possible to select for resistance against patent strongylid infection, although a possible antagonistic relation between the infection with strongylids and RAO must be considered.

### Manufacturers' addresses

- <sup>1</sup> John C. Pezzullo, Georgetown University, Washington D.C., USA
- <sup>2</sup> SAS Institute Inc., Cary, North Carolina, USA
- <sup>3</sup> Hasstedt, S.J. 1994. PAP: Pedigree Analysis Package, Rev. 4.0, Department of Human Genetics, University of Utah, Salt Lake City, USA
- <sup>4</sup> Gill, P.E., W. Murray, M.A. Saunders, and M.H. Wright. 1986. NPSOL: A Fortran package for nonlinear programming. Technical Report SOL 86-2, Stanford University, Stanford, CA
- <sup>5</sup> Janss, L.L.G. 2008. "iBay manual version 1.44". Janss Bio-statistics, Leiden, Netherlands

### References

- Ardeleanu D., Neacsu M. G., Pivoda C. A., Neacsu C. and Ardeleanu I. C. (2008) Antiparasitilic protection in sheep farms. *Lucruri stiintifice Zootehnie si Biotehnologii* 41, 721-726.
- Barnes K. C., Grant A. V. and Gao P. (2005) A review of the genetic epidemiology of resistance to parasitic disease and atopic asthma: common variants for common phenotypes? *Curr. Opin. Allerg. Clin. Immunol.* 5, 379-385

- Bauer C. (2006) Nachweis von Parasitenstadien im Kot. In: Veterinärmedizinische Parasitologie, 6th edn., Ed T. Schnieder, Parey, Stuttgart. pp 88-96
- Blaser K. (2008) Immunological principles of allergen-specific immune therapy. *HNO* 55, 759-763
- Bründler P., Frey C. F., Gottstein B., Nussbaumer P., Neuhaus S. and Gerber V. (2011) Lower shedding of strongylid eggs by Warmblood horses with recurrent airway obstruction compared to unrelated healthy horses. *Vet. J.* 9, Epub ahead of print (doi: 10.1016/j.tvjl.2010.12.029).
- da Rocha F. A. (2006) Protective role of helminthiasis in the development of autoimmune diseases. *Clin. Dev. Immunol.* 13, 159-162
- Davies G., Stear M. J., Benothman M., Abuagob O., Kerr A., Mitchell S. and Bishop S. C. (2006) Quantitative trait loci associated with parasitic infection in Scottish blackface sheep. *Heredity* 96, 252-258
- Gerber V., Lindberg A., Berney C. and Robinson N. E. (2004) Airway mucus in recurrent airway obstruction-short-term response to environmental challenge. *J. Vet. Intern. Med.* 18, 92-97
- Gerber V., Baleri D., Klukowska-Rötzler J., Swinburne J. E. and Dolf G. (2009) Mixed inheritance of equine recurrent airway obstruction. *J. Vet. Intern. Med.* 23, 626-630
- Giles C. J., Urquhart K. A. and Longstaffe J. A. (1985) Larval cyathostomiasis (immature trichonema-induced enteropathy): A report of 15 clinical cases. *Equine Vet. J.* 18, 196-201
- Hershey G. K. K., Friedrich M. F., Esswein L. A., Thomas M. L. and Chatila T. A. (1997) The association of atopy with a gain-of-function mutation in the alpha subunit of the interleukin-4 receptor. *N. Engl. J. Med.* 337, 1720-1725
- Ishikawa Y., Yoshimoto T. and Nakanishi K. (2006) Contribution of IL-18-induced innate T cell activation to airway inflammation with mucus hypersecretion and airway hyperresponsiveness. *Int. Immunol.* 18, 847-855
- Jost U., Klukowska-Rötzler J., Dolf G., Swinburne J.E., Ramseyer, Bugno M., Burger D., Blott S. and Gerber V. (2007) A region on equine chromosome 13 is linked to recurrent airway obstruction in horses. *Equine Vet. J.* 39, 236-241
- Krecek R. C., Reinecke R. K. and Horak I. G. (1989) Internal parasites of horses on mixed grassveld and bushveld in Transvaal, Republic of South Africa. *Vet. Parasitol.* 34, 135-143
- Love S. and Duncan J. L. (1992) The development of naturally acquired cyathostome infection in ponies. *Vet. Parasitol.* 44, 127-142.
- Love S., Murphy D. and Mellor D. (1999) Pathogenicity of cyathostome infection. *Vet. Parasitol.* 85, 113-122
- Lynch N. R., Hagel I. A., Palenque M. E., Di Prisco M. C., Esudero J. E., Corao L. A., Sandia J. A., Ferreira L. J., Botto C., Perez M. and Le Souef P. N. (1998) Relationship between helminthic infection and IgE response in atopic and nonatopic children in a tropical environment. *J. Allergy. Clin. Immunol.* 217-221
- Marti E., Gerber H., Essich G., Oulehla J. and Lazary S. (1991) The genetic basis of equine allergic diseases 1. Chronic hypersensitivity bronchitis. *Equine Vet. J.* 23, 457-460
- Martin J. (2001) Human asthma and chronic obstructive pulmonary disease (COPD). International workshop on equine chronic airway disease, Michigan State University, 16-18 June 2000. *Equine Vet. J.* 33, 5-19
- McKay D. M. (2006) The beneficial helminth parasite, *Parasitology* 132, 1-12.
- Meier A. and Hertzberg H. (2005) Strongyliden beim Pferd. II. Vorkommen von Anthelminthika-Resistenzen in der Schweiz. *Schweiz. Arch. Tierheilk.* 9, 389-396
- Nakae S., Komiyama Y., Yokoyama H., Nambu A., Umeda M., Iwase M., Homma I., Sudo K., Horai R., Asano M. and Iwakura Y. (2003) IL-1 is required for allergen-specific Th2 cell activation and the development of airway hypersensitivity response. *Int. Immunol.* 15, 483-490
- Neuhaus S., Bruendler P., Frey C. F., Gottstein B., Doherr M. G. and Gerber V. (2010) Increased parasite resistance and recurrent airway obstruction in horses of a high-prevalence family. *J. Vet. Intern. Med.* 24, 407-413
- Ober C., Tsalenko A., Parry R. and Cox N. (2000) A second-generation genomewide screen for asthma. Susceptibility alleles in a founder population. *Am. J. Hum. Genet.* 67, 1154-1162
- Ogbourne C. P. (1976) The prevalence, relative abundance and site distribution of nematodes of the subfamily Cyathostominae in horses killed in Britain. *J. Helminthol.* 50, 203-214
- Paterson S., Wilson K. and Pemberton J. M. (1998) Major histocompatibility complex variation associated with juvenile survival and parasite resistance in a large unmanaged ungulate population (*Ovis aries* L.). *Proc. Natl. Acad. Sci. U S A.* 95, 3714-3719
- Ramseyer A., Gaillard C., Burger D., Straub R., Jost U., Boog C., Marti E. and Gerber V. (2007) Effects of genetic and environmental factors on Horse Owner Assessed Respiratory Signs Index (HOARSI). *J. Vet. Intern. Med.* 21, 149-156
- Reinemeyer C. R., Smith S. A., Gabel A. A. and Herd R. P. (1984) The prevalence and intensity of internal parasites of horses in the U.S.A.. *Vet. Parasitol.* 15, 75-83
- Robinson N. E. (2001) International workshop on equine chronic airway disease, Michigan State University, 16-18 June 2000. *Equine Vet. J.* 33, 5-19
- Sasaki Y., Yoshimoto T., Maruyama H., Tegoshi T., Ohta N., Arizono N. and Nakanishi K. (2005) IL-18 with IL-2 protects against *Strongyloides venezuelensis* infection by activating mucosal mast cell-dependent type 2 innate immunity. *J. Exp. Med.* 202, 607-616
- Scales H. E., Ierna M. X. and Lawrence C. E. (2007) The role of IL-4, IL-13 and IL-4Ralpha in the development of protective and pathological responses to *Trichinella spiralis*. *Parasite Immunol.* 29, 81-91
- Seaton G., Hernandez J., Grunchev J. A., White I., Allen J., De Koning D. J., Wei W., Berry D., Haley C. and Knott S. (2006) GridQTL: A Grid Portal for QTL Mapping of Compute Intensive Datasets. Proceedings of the 8th World Congress on Genetics Applied to Livestock Production, August 13-18, 2006. Belo Horizonte, Brazil
- Strachan D. P. (1989) Hay fever, hygiene, and household size. *Br. Med. J.* 299, 1259-1260
- Swinburne J. E., Bogle H., Klukowska-Rötzler J., Drögemüller M., Leeb T., Temperton E., Dolf G. and Gerber V. (2009) A whole-genome scan for recurrent airway obstruction in Warmblood sport horses indicates two positional candidate regions. *Mamm. Genome.* 20, 504-515
- Titus, R.G., Sherry, B. and Cerami, A. (1991) The involvement of TNF, IL-1 and IL-6 in the immune response to protozoan parasites. *Immunol. Today* 12, 13-16.
- von Samson-Himmelstjerna G. (2006) Helminthosen der Equiden. In: Schnieder T, ed. *Veterinärmedizinische Parasitologie*. 6th edition. Parey Stuttgart, 303-347

Päivi Nussbaumer Schleuniger  
Equine Clinic  
Department of Clinical Veterinary Medicine  
Vetsuisse-Faculty  
University of Berne  
Länggassstrasse 124  
3012 Berne  
Switzerland  
paeivi.nussbaumer@haras.admin.ch